* Plate out isolates on LB, 2 isolates / plate
* Wash chips – 2 mL 10% bleach, 0.1 N NaOH, autoclaved dH2O. Then rinse in water, then NaOH, then water.
* Start overnight in 1 mL LB broth in deep well plates, 37 C, 1000 rpm
* The next day, flow through 1 mL H2O.
* Use excel to generate plate layout
  + See protocol
    - blanks on outer edge and 20 more randomized
    - Blanks get 100 uL LB
    - Condition 1 gets 90 uL LB
    - Condition 2 gets 89 uL LB
    - Condition 3 gets 88 uL LB
    - 40 mL LB total
    - Pipette LB with a high volume tube
    - 1 uL of amox or clav acid
    - 10 uL cells / well
    - Randomized
  + Copy into dispense list
* Dispense media
  + Can use same chip for clavulanic acid and media
  + Washed tube by flowing through 1000 ul of each cleaning
* Make antibiotics and cells
  + Make 25 mg/mL amoxicillin (5 mg in 200 uL DMSO)
    - Dilute 100 uL in 400 uL LB to get 5 mg/mL, 500 uL total (need 192, load 450)
  + Make 12.5 mg/mL clavulanic acid (5 mg / 400 uL H2O)
    - Sterile filter
    - Dilute 50 uL in 200 uL LB to get 2.5 ug/mL, 250 uL total (need 96, load 250)
  + Dilute isolates
    - Mix 180 uL LB with 20 uL cells
    - Measure OD (blank and multiply by 10)
    - Correct to OD 1
    - Dilute resulting culture 50 uL in 350 uL LB to get 1 x 10^8 cells / mL
    - 400 uL for each isolate (need 120, load 350)
* Dispense everything else
  + Prime 30 uL and predispense 12
* Ethanol lid for lid lifter
* Measurements in 10 minute increments for 24 hours at 30 C on the spark with the lid lifter
* Wash chips